

**IDENTIFICATION OF COMMONLY USED MEDICINAL PLANTS IN  
KANGKAR PULAI AND THEIR THERAPEUTIC  
EVALUATION AS ANTI-DIABETIC**

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**UNIVERSITI TEKNOLOGI MALAYSIA**

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*Specially dedicated goes to:*

*My dear parents*

*My wife*

*My siblings*

*My friends*

*For their love, understanding and support through my endeavour*

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## ABSTRACT

Medicinal plants have been used as traditional medicines for treatment of diseases such as diabetes mellitus. However, scientific information to support the claims of herbal medicine practitioners based on Use Value (UV) and Informant Consensus Factor (ICF) is largely unexplored. The aim of this study is to screen common medicinal plants and to evaluate their traditional use through an ethnopharmacological survey at Kangkar Pulai, Johor, Malaysia. Data were collected from the community through open interviews, determining the relative importance of the species surveyed and calculating the UV and ICF in relation to the medicinal plant uses. A total of 38 plant species belongs to 28 families were documented. Based on the results of ethnopharmacological survey, *Orthosiphon stamineus* Benth and *Momordica charantia* L recorded the highest Use Values (0.32 and 0.24) to treat diabetes. The results of antioxidant tests revealed that the total phenolic content (TPC) of *O. stamineus* is  $71.70 \pm 0.85$  mg (GAE)/g and total flavonoid content (TFC) is  $44.71 \pm 0.75$  mg (CE)/g. Further study was carried out to examine the effects of oral administration of *O. stamineus* extract in the treatment of diabetes in normal and alloxan-induced diabetic rats. Forty rats were divided into four groups of 10 each. Group A (control) consisted of normal rats receiving 2 mL (10 mL/kg bwt) of normal saline on daily basis, whereas group B consisted of diabetic rats treated with 1 mL (120 mg/kg bwt) of *O. stamineus* extract. Group C consisted of diabetic rats treated with 1 mL (150 mg/kg bwt) of Metformin. Group D consisted of untreated diabetic rats acted as negative control. Group B, C and D were injected intraperitoneally with alloxan (150 mg /kg bwt). Diabetic group B rats treated with *O. stamineus* extract showed significantly ( $p < 0.05$ ) low blood glucose level compared to group D (untreated diabetic rats). Similarly diabetic group B rats consumed significantly lower daily food and water intake at significant level  $p < 0.05$  compared to group D (untreated diabetic rats). Diabetic group B rats treated with *O. stamineus* extract showed significantly higher body weight at significant level  $p < 0.05$  compared to group D (untreated diabetic rats). Diabetic group B rats treated with *O. stamineus* extract showed lower serum total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL-C) and higher high-density lipoprotein cholesterol (HDL-C) at significant value ( $p < 0.05$ ) compared with group D (untreated diabetic rats). A significant decrease of total protein, albumin and creatinine was observed in the plasma of group B rats after being treated with *O. stamineus* extract compared with group D. Additionally, immunohistochemistry results showed that the levels of heat shock protein 70 (HSP70) and inducible nitric oxide synthase (iNOS) of group B diabetic rats were brought back to near normal range after being treated with *O. stamineus* extract at significant value ( $p < 0.05$ ) compared with group D (untreated diabetic rats). In conclusion, *O. stamineus* extract exhibited antidiabetic activity in alloxan-induced diabetic rats. Thus, the present findings also support the potential use of *O. stamineus* extract as a remedy for hyperglycemia

## ABSTRAK

Tumbuhan perubatan telah digunakan sebagai ubat-ubatan tradisional untuk merawat penyakit seperti diabetes mellitus. Walaubagaimanapun, maklumat saintifik berdasarkan Nilai Gunaan (UV) dan Persetujuan Pemberi Maklumat (ICF) bagi menyokong dakwaan pengamal perubatan tumbuhan masih belum diterokai. Tujuan kajian ini adalah untuk meninjau tumbuhan perubatan biasa dan untuk menilai penggunaan tradisional mereka melalui kaji selidik ethnopharmacology di Kangkar Pulai, Johor, Malaysia. Data-data telah didapati dari masyarakat melalui temuduga secara langsung, menentukan perbezaan kepentingan spesies yang ditinjau dan mengira Faktor Persetujuan Pemberi Maklumat (ICF) dan Nilai Gunaan (UV) berhubung penggunaan tumbuhan perubatan. Sebanyak 38 spesies tumbuhan yang dimiliki oleh 28 keluarga telah didokumenkan. Berdasarkan hasil kajian ethnopharmacological, *Orthosiphon stamineus* Benth dan *Momordica charantia* L mencatatkan nilai penggunaan yang tertinggi sebagai rawatan penyakit diabetes dengan nilai penggunaan, 0.32 dan 0.24, masing-masing. Keputusan ujian antioksidan menunjukkan ekstrak *O. stamineus* mengandungi jumlah kandungan fenol (TPC)  $71.70 \pm 0.85$  mg (GAE) / g dan jumlah kandungan flavonoid (TFC)  $44.71 \pm 0.75$  mg (CE)/g. Kajian lanjut telah dijalankan untuk mengkaji kesan pengambilan ekstrak *O. stamineus* melalui mulut terhadap rawatan penyakit diabetes pada tikus normal dan diabetes yang disuntik dengan alloxan. Empat puluh ekor tikus telah dibahagikan kepada empat kumpulan dengan 10 ekor bagi setiap kumpulan. Kumpulan A terdiri daripada tikus-tikus normal yang menerima 2 mL (10 ml/kg bwt) larutan garam biasa (kawalan) pada setiap hari, manakala kumpulan B terdiri daripada tikus-tikus diabetes yang dirawat dengan 1 mL (120 mg/ kg bwt) ekstrak *O. stamineus*. Kumpulan C pula terdiri daripada tikus-tikus diabetes yang dirawat dengan 1 mL (150 mg/kg bwt) Metformin. Manakala kumpulan D terdiri daripada tikus-tikus diabetes yang tidak dirawat bertindak sebagai kumpulan kawalan negatif. Kumpulan B, C dan D telah disuntik intraperitoneally dengan Alloxan (150 mg / kg bwt). Tikus-tikus diabetes kumpulan B yang dirawat dengan ekstrak *O. stamineus* menunjukkan dengan ketara ( $p < 0.05$ ) tahap glukosa darah yang rendah berbanding dengan kumpulan D (tikus-tikus diabetes yang tidak dirawat). Demikian juga tikus-tikus kumpulan B menunjukkan dengan ketara ( $p < 0.05$ ), pengambilan makanan harian dan air yang lebih rendah berbanding dengan kumpulan D (tikus diabetes yang tidak dirawat). Mereka (kumpulan B) juga menunjukkan dengan ketara ( $p < 0.05$ ) berat badan yang lebih tinggi, lebih rendah jumlah kolesterol, trigliserida, kolesterol lipoprotein ketumpatan rendah dan lebih tinggi kolesterol lipoprotein ketumpatan tinggi dalam serum berbanding dengan kumpulan D (tikus diabetes tidak dirawat). Penurunan yang ketara didapati bagi jumlah protein, albumin dan kreatinin dalam plasma tikus-tikus kumpulan B selepas rawatan dengan ekstrak *O. stamineus*. Selain itu, keputusan immunohistokimia menunjukkan tahap protein kejuthaba 70 (HSP70) dan inducible nitrik oksida sintase (iNOS) telah dikembalikan kepada julat hampir biasa selepas rawatan dengan ekstrak *O. stamineus* dalam Kumpulan B pada nilai signifikan ( $p < 0.05$ ) berbanding dengan kumpulan D (tikus-tikus diabetes yang tidak dirawat). Kesimpulannya, ekstrak *O. stamineus* menunjukkan aktiviti anti-diabetes pada tikus-tikus diabetes yang disuntik Alloxan. Maka oleh kerana itu, penemuan ini juga menyokong potensi penggunaan ekstrak *O. stamineus* sebagai ubat untuk hiperglisemia.

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## LIST OF ABBREVIATIONS

ADP	- Adenosine diphosphate
AGEs	- Advanced glycosylation end products
AIDS	- Acquired immunodeficiency syndrome
AlCl <sub>3</sub>	- Aluminium chloride
ATP	- adenosine triphosphate
CAE	- catechin equivalents
cNOS	- Constitutive NOS, including eNOS and nNOS
DAB	- 3,3' - diaminobenzidine
DMEM	- Dulbecco's Modified Eagle's Medium
DMSO	- dimethyl sulphoxide
DNA	- Deoxyribonucleic acid
DPPH	- 1,1-Diphenyl-2-picrylhydrazyl
DPX	- Dextrane plasterized xylene
EDTA	- Ethylenediaminetetraacetic acid
EMEM	- Eagle's Minimum Essential Medium
eNOS	- Endothelial NOS
GAE	- Gallic Acid Equivalents
GLUT2	- glucose transporter 2
HDL	- High density lipoprotein
HIV	- Human immunodeficiency virus
HSPs	- Heat shock proteins
I	- Type one
ICF/Fic	- informant consensus factor
IDDM	- Insulin-dependent diabetes mellitus
IGT	- impaired glucose tolerance

II	- Type two
IKK- $\beta$	- inhibitory protein $\kappa$ B kinase $\beta$
iNOS	- inducible Nitric Oxide Synthase
IPNI	- International Plant Names Index
IR	- Insulin receptor
IRS	- insulin receptor substrate
LDL	- Low-density lipoprotein
LMIC	- Lower middle income countries
MafA	- musculoaponeurotic fibrosarcoma protein A
MAPK	- mitogen-activated protein kinase
MODY	- maturity-onset diabetes in youth
mRNA	- Messenger RNA
MTT	- methyl tetrazolium
N	- number of informants
Na <sub>2</sub> CO <sub>3</sub>	- Sodium carbonate
NaNO <sub>2</sub>	- Sodium nitrite
NaOH	- Sodium hydroxide
NEFAs	- non-esterifies fatty acids
NF $\kappa$ B	- Nuclear transcription factor $\kappa$ B
NF-kB	- nuclear factor-kB
NIDDM	- Non-insulin-dependent diabetes mellitus
nm	- Nanometers
nNOS	- Neuronal NOS
NO	- Nitric oxide
Nt	- number of taxa used
Nur	- number of use reports per each category
O <sub>2</sub>	- superoxide anion
OGTT	- oral glucose tolerance test
PCV	- Packed cell volume
PDX-1	- Pancreatic and duodenal homeobox 1
PI3K	- phosphatidylinositol-3-OH kinase
PKB	- Akt/ protein kinase B
PKC	- Protein kinase C



R	- Reagent
RBC	- red blood cell
ROS	- Reactive oxygen species
STZ	- Streptozotocin
T2DM	- Type 2 Diabetes Mellitus
TG	- Triglyceride
U	- number of uses per species
UCP-2	- uncoupling protein-2
UN	- United Nations
UNESCO	- United Nations Educational, Scientific and Cultural Organization
US\$	- United States dollar
USA	- United States of America
UV	- Use value
UV light	- Ultraviolet
V	- Volume
VLDL	- Very-low-density lipoprotein
W	- Weight
WBC	- white blood cell
WHO	- World Health Organization

**LIST OF SYMBOLS**

G	-	Gram
Kg	-	Kilogram
Dl	-	Deciliter
ml	-	Milliliters
$\mu$ L	-	Microliters
M	-	Molarity
mm	-	Millimeters
$^{\circ}$ C	-	Celsius
$^{\circ}$ F	-	Fahrenheit
Mg	-	Milligram
Ng	-	Nanograms
Pg	-	Pictogram

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## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of Research**

In the last few decades, there have been many studies on ethno-medicine (Krippner, 2003; Williams, 2006). Ethno-medicine is the study of traditional medical practice. It is related to the cultural interpretation of health, diseases, and illnesses. It basically addresses matters concerning healthcare seeking and healing processes and practices (Krippner, 2003; Williams, 2006). Ethno-medicine is a highly complicated multi-disciplinary system that explores the use of plants, spirituality, and the natural environment, which has been the main source of treatment and healing for many people over the years (Lowe *et al.*, 2001).

With the emergence of the discipline, research in the field of ethno-medicine has significantly contributed towards a better understanding of traditional subsistence, as well as medical knowledge and practice. A vast amount of literature on ethno-medicine has been motivated by the increasing awareness about the consequences of forced displacement and acculturation of the local people, the recognition of indigenous health practices and concepts to maintain ethnic identity, and the search for new medical cures and technologies (Williams, 2006).

The World Health Organization, (2003) reported that ethno-medicine has sustained its popularity in the developing world and its use is fast gaining roots in the developed world as well. The traditional herbal preparations of China accounts for 30–50% of the total consumption of medicines (WHO, 2003). In African countries, such as Ghana, Nigeria, Zambia, and Mali, herbal medicine is used to treat 60% of the children affected with malaria. In London, San Francisco, and South Africa, 70% of people suffering from HIV/AIDS are also reported to use herbal medicines at a cost of more than US\$60 billion (WHO, 2003).

The World Health Organization has defined traditional medicine as the body of knowledge and practices used to recognize, prevent, and/or reduce some physical, mental, or social diseases that may rely on past experience and observation handed down from one generation to another in either verbal or written form (WHO, 1999a). In developing countries, complementary traditional medicines are often used. According to estimates by the World Health Organization, almost 80% of people living in rural areas in many countries are looked after by traditional medical practitioners. Another approximation is that about half of industrialized countries reportedly use traditional medicines regularly. Modern pharmaceutical agents, including many plant-derived therapeutic agents, are also supported by traditional herbal plants (Alzweiri *et al.*, 2011; Alsarhan *et al.*, 2012)

Over the years, people have used different materials from nature to improve their health and to treat their diseases. For this purpose, various substances were derived from animals, plants, and mineral resources in areas where people lived, as well as from very remote places (Ghorbani *et al.*, 2006). Nature has been the main source of medicinal agents for thousands of years. A large number of natural drugs have been isolated from natural sources to treat various diseases by keeping in mind the use of traditional medicine. This type of plant-based traditional medicine continues to play an important role in the practice of healthcare. About 80% of the world population uses conventional medicine prepared from various traditional medicinal plants as their primary healthcare (Ghorbani *et al.*, 2006).

Throughout the years, traditional medicines have been proven to be an invaluable source and guidance for screening of drugs. Many important and famous modern drugs, such as digitoxin, reserpine, tubocurarine, ephedrine, ergometrine, atropine, vinblastine, and aspirin, were discovered on the basis of traditional folk medicine (Anyinam, 1995). In many parts of the world, medicinal herbs are considered an important element of the medical system of the indigenous people, and these resources are also an essential part of the traditional knowledge of the culture (Ghorbani *et al.*, 2006).

Although reports have revealed improvements in the quality of life and life prolongation in diabetic patients after using traditional medicines, the biological activity of such medicines has not been documented (Bailey and Day, 1989). In general, with regard to traditional medicines in Asian countries, many herbal medicines are put together to make a multi-herbal formula to enhance its function (Bailey and Day, 1989).

Herbal medicines are still popular despite the abundance of modern medicine for cultural and historical reasons. Information is also available about the uses of herbal plants for the treatment of different diseases all over the world. However, there is a need to further investigate, correlate, and document these plants (Alzweiri *et al.*, 2011; Alsarhan *et al.*, 2012).

Diabetes belongs to a group of metabolic diseases having high blood sugar (glucose) levels, which are the result of defects in insulin secretion or action, or even both. Insufficient action of insulin results in increased blood-glucose concentration. The most common metabolic disorder (hyperglycemia) in the world is diabetes mellitus (Tierney *et al.*, 2002).

In the progress of diabetes, numerous pathogenic events are involved, ranging from autoimmune damage of  $\beta$ -cells in the pancreas resulting in insulin insufficiency

to abnormalities that results in resistance to the insulin action (Alberti and Zimmet, 1998; American Diabetes Association, 2013).

The cause for defects in carbohydrate, lipid, and protein metabolism in diabetes is incomplete action of insulin on the specific tissues. The reason for deficient insulin action is inadequate insulin excretion or diminishing tissue responses to insulin action at one or more points in the complicated paths of hormone activity. Deficiency of insulin production and defects in insulin activity frequently exist in the same patient, and it is often indistinct which anomaly, if either alone, is the main reason for the hyperglycemia (Gavin *et al.*, 1997; American Diabetes Association, 2013).

Hyperglycemia symptoms include polyuria, polydipsia, weight loss, polyphagia and blurred vision. Weakness of growth and exposure to certain infections may induce chronic hyperglycemia. The life-threatening and acute after-effects of uncontrolled diabetes is hyperglycemia with ketoacidosis or non-ketotic hyperosmolar disease (Gavin *et al.*, 1997; American Diabetes Association, 2013).

The complications of diabetes include retinopathy or vision loss, and nephropathy. This leads to renal insufficiency and peripheral neuropathy with the risk of foot sores, amputations, and Charcot joints. Autonomic neuropathy causes gastrointestinal, genitourinary, and cardiovascular diseases, and even sexual dysfunction. Those patients with diabetes complications have increased prevalence of peripheral arterial and cardiovascular diseases. As well as this, diabetes patients have hypertension and abnormalities of lipoprotein metabolism (American Diabetes Association, 2013).

Insulin is a hormone that helps glucose enter the cells in the body to provide energy. Symptoms that appear in patients are frequent urination, lethargy, excessive thirst, and loss of appetite. Diabetes can be treated with dietary changes, timely

medication, and, in some cases, by administering insulin injections on a daily basis. This depends on the type and severity of the problem (Bhikha and Glynn, 2013).

Type 1 diabetes is normally treated by using insulin, doing exercise, and through maintaining a diabetic diet. Type 2 diabetes is initially treated by weight loss measures, maintaining a diabetic regimen, and exercise. When these measures do not reduce the problem of raised blood sugars, then oral medications are used. In the case of failure of oral medication, insulin medications are considered. Natural herbs are also traditionally used to treat type 2 diabetes mellitus (Thomas *et al.*, 2004).

Diabetes can be treated through diet, exercise, oral hypoglycemic agents, and insulin. Today, a synthetic drug is available and is considered as an anti-diabetic agent, but it is expensive and produces serious side effects. In addition to the currently available therapeutic options, numerous herbal medicines are mentioned for treating diabetes mellitus. Generally, medicinal plants are advantageous due to the lack of side effects (Ayodhya *et al.*, 2010; Elavarasi *et al.*, 2013).

## **1.2 Problem Statements**

Medicinal plants continue to play an important role in the treatment of diabetes, particularly in developing countries where most people have limited resources and do not have access to modern treatment. The increase in demand in industrially developed countries to use alternative approaches to treat diabetes, such as plant-based medicines, is also due to the side effects associated with the use of insulin and oral hypoglycemic agents (Anumsima, 2011).



Testing of the biological activity of medicinal plants for the treatment of diabetes mellitus based on ethnopharmacological studies may be hindered by inaccurate information collected from indigenous people regarding the use of medicinal plants, the weak points in these studies that might be overcome in the design of studies and interpretation of data. In Malaysia, there are about 12,000 kinds of flowering plants, of which about 1300 have been documented as medicinal plants, and only about 100 have been extensively studied for their medicinal effect (Anumsima, 2011).

Even with the several medicinal plants documented by ethnopharmacological studies for the treatment of diabetes, there are weak points in the determination of the use value for each plant and the informant consensus factor, because the use value explains the number of medicinal plants that have highest use value for the treatment of diabetes mellitus. Additionally, this knowledge is required to prove the validity of the claimed medicinal uses as recommended by traditional healers for the treatment of diseases, including diabetes. In order to overcome this situation, systematic research is needed to identify inexpensive, harmless, and effective anti-diabetic drugs.

Many studies have been conducted based on traditional knowledge and phytochemical analysis, but the combination of ethnopharmacological study and animal research is more reliable. Keeping in view the importance of using medicinal plants for the treatment of diabetes, this study aimed to identify and document the major medicinal plants being used by people in the Kangkar Pulai area, Johor, Malaysia, for treating human diseases. This study investigated the informant consensus factor and the use value of the plants in Kangkar Pulai area for treating diabetes and other diseases. Moreover, this ethno-medicine survey used certain equations to determine the types of medicinal plants and diseases that are treated by these plants. This study used specific biomarkers (HSP70 and iONS) to evaluate the effect of selected plants on the treatment of diabetic rats induced by alloxan.

### **1.3 Objectives of the Study**

The main objective of this study was to investigate the anti-diabetic properties of one of the Malaysian herbs based on the ethnopharmacological survey combined with an animal model study. The objectives can be sub-classified as follows:

1. To determine the use value and informant consensus factor of traditional medicinal plants used for the treatment of diabetes based on an ethnopharmacological survey in Kangkar Pulai, Johor, Malaysia.
2. To determine the total phenol and flavonoid content, as well as the antioxidant activity, of selected medicinal plants.
3. To investigate the anti-diabetic properties of selected medicinal plant in a rat model of type 1 diabetes.

### **1.4 Scope of the Study**

The following scopes of the study were identified:

- 1.4.1 To determine the informant consensus factor and use value of local herbs, particularly for diabetic treatment based on open interviews of 25 volunteers in Kangkar Pulai, Johor, Malaysia.
- 1.4.2 To determine the total phenol and flavonoid content, as well as the DPPH values of phytochemicals in selected anti-diabetic plants based on spectrophotometric method.

- 1.4.3 To evaluate the anti-diabetic properties of selected anti-diabetic plant based on glucose level, food intake, water intake, hematological parameters, and immunohistological analysis of the pancreas and liver of Alloxan-induced diabetic rats.

## REFERENCES

- AbuKhader, M. (2012). The Effect of Route of Administration in Thymoquinone Toxicity in Male and Female Rats. *Indian journal of pharmaceutical sciences*, 74(3), 195.
- Adebayo, J. O., Adesokan, A. A., Olatunji, L. A., Buoro, D. O. and Soladoye, A. O. (2005). Effect of Ethanolic Extract of Bougainvillea Spectabilis Leaves on Haematological and Serum Lipid Variables in Rats. *Biokemistri* 2005;17: 45-50.
- Adelson, N. (1993). *Being Alive Well: Indigenous Belief as Opposition Among the Whapmagoostui Cree*. PhD Dissertation, McGill University. Canada
- Adenan, M. (2003). Malaysian Herbs and Herbal Products. *A Two and Half Day Course of Herbal and Phytochemical Processing. CEPP Short Course Notes* (Chemical Engineering Pilot Plant, Universiti Teknologi Malaysia, Malaysia).
- Aja, E. J. Nwafor, A. U. Ibiam, O. U. Orji, Ezeani, N. and Nwali, a. B. U. (2013). Evaluation of Anti-diabetic and Liver Enzymes Activity of Aqueous Extracts of *Moringa oleifera* and *Bridelia ferruginea* Leaves in Alloxan Induced Diabetic Albino Rats. *International Journal of Biochemistry Research & Review*, 3(3), 248-258.
- Ajao, S. M., Olayaki, L. A., Oshiba, O., Jimoh, R. O., Jimoh, S. A., Olawepo, A. (2009). Comparative Study of the Hypoglycemic Effects of Coconut Water Extract of Picralima Nitida Seeds (Apocynaceae) and Daonil in alloxan-Induced Diabetic Albino Rats. *African Journal of Biotechnology*, 8(4).

- Akinola, O., Gabriel, M., Suleiman, A. A., & Olorunsogbon, F. (2012). Treatment of Alloxan-Induced Diabetic Rats with Metformin or Glitazones is Associated with Amelioration of Hyperglycaemia and Neuroprotection. *The Open Diabetes Journal*, 5, 8.
- Akowuah, G. A., Zhari, I., Norhayati, I., Sadikun, A. and Khamsah, S. M. (2004). Sinensetin, Eupatorin, 3'-hydroxy-5, 6, 7, 4'-tetramethoxyflavone and rosmarinic Acid Contents and Antioxidative Effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*, 87(4), 559-566.
- Al-Jarrah, M., Matalka, I., Al Aseri, H., Mohtaseb, A., Smirnova, I. V., Novikova, L. (2010). Exercise Training Prevents Endometrial Hyperplasia and Biomarkers for Endometrial Cancer in Rat Model of Type 1 Diabetes. *Journal of clinical medicine research*, 2(5), 207.
- Al-khatib, A. (2013). Co-Expression of Inos and Hsp70 in Diabetes Type 1 Makes a Rational Hypothesis to Explain the Diabetic Neuropathy. *European Scientific Journal*, 9(3).
- Al-Shamaony, L., Al-Khazraji, S. M. and Twaij, H. A. (1994). Hypoglycaemic Effect of *Artemisia Herba Alba*. II. Effect of a Valuable Extract on Some Blood Parameters in Diabetic Animals. *Journal of Ethnopharmacology*, 43(3), 167-171.
- Alberti, K. G. M. M. and Zimmet, P. (1998). Definition, Diagnosis and Classification of Diabetes Mellitus and its Complications. Part 1: Diagnosis and Classification of Diabetes Mellitus. Provisional Report of a WHO Consultation. *Diabetic medicine*, 15(7), 539-553.
- Allain, C. C., Poon, L. S., Chan, C. S., Richmond, W. and Fu, P. C. (1974). Enzymatic Determination of Total Serum Cholesterol. *Clinical Chemistry*, 20(4), 470-475.
- Almdal, T., Scharling, H., Jensen, J. S. and Vestergaard, H. (2004). The Independent Effect of Type 2 Diabetes Mellitus on Ischemic Heart Disease, Stroke, and Death: A Population-Based Study of 13 000 Men and Women With 20 Years of Follow-Up. *Archives of Internal Medicine*, 164(13), 1422.
- Alsarhan, A., Sultana, N., Kadir, M. and Aburjai, T. (2012). Ethnopharmacological Survey of Medicinal Plants in Malaysia, the Kangkar Pulai Region. *International Journal of Pharmacology*, 8, 679-686.

- Alsarhan, A., Mansi, K., Aburjai, T., Al-Khatib, A., Alzweiri, M. and Sultana, N. (2013). HSP70 and Inos Biomarkers in Evaluating the Healing Properties of Rubia Tinctorum. *European Scientific Journal*, 9(15), 1857-7881.
- Alsarhan, A., Sultana, N., Al-Khatib, A. and Kadir, M. R. A. (2014). Review on Some Malaysian Traditional Medicinal Plants with Therapeutic Properties. *Journal of Basic and Applied Sciences*, 10, 149-159.
- Alshawsh, M. A., Abdulla, M. A., Ismail, S., Amin, Z. A., Qader, S. W., Hadi, H. A., & Harmal, N. S. (2012). Free Radical Scavenging, Antimicrobial and Immunomodulatory Activities of Orthosiphon Stamineus. *Molecules*, 17(5), 5385-5395.
- Alzweiri, M., Sarhan, A. A., Mansi, K., Hudaib, M. and Aburjai, T. (2011). Ethnopharmacological Survey of Medicinal Herbs in Jordan, the Northern Badia Region. *Journal of Ethnopharmacology*, 137(1), 27-35.
- American Diabetes Association, (2013). Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care*, 36:Supplement 1 S7-S74.
- Amos, A. F., McCarty, D. J. and Zimmet, P. (1997). The Rising Global Burden of Diabetes and its Complications: Estimates and Projections to the Year 2010. *Diabetic Medicine*, 14(S5), S7-S85.
- Anjaneyulu, M. and Chopra, K. (2005). Diltiazem Attenuates Oxidative Stress in Diabetic Rats. *Renal Failure*, 27(3), 335-344.
- Ansarullah, B. B., Patel, V. and Ramachandran, A. (2012). Improved Glucoregulation, Insulin Resistance and Leptin Levels by a Polyherbal Drug in High Fat Diet and Low Dose Streptozotocin Type 2 Diabetes Model. *Diabetologia Croatica*, 41(1),3.
- Anumsima ahmad barkat. (2011). Optimization of Aqueous Extraction Process for  $\beta$ -Glucosidase Inhibitory Compounds from Selected Malaysian Herbs, Master Thesis , Kuliyah of Engineering , International Islamic University Malaysia.
- Anyinam, C, (1995). Ecology and Ethno-Medicine: Exploring Links between Current Environmental Crisis and Indigenous Medical Practices. *Social Science and Medicine*, 40(3): 321–329.
- Ashraf, H., Heidari, R., Nejati, V., & Ilkhanipoor, M. (2012). Aqueous Extract of Berberis Integerrima Root Improves Renal Dysfunction in Streptozotocin Induced Diabetic Rats. *Avicenna Journal of Phytomedicine*, 3(1), 82-90.

- Arafat, O., Tham, S., Sadikun, A., Zhari, I., Haughton, P. and Asmawi, M. (2008). Studies on Diuretic and Hypouricemic Effects of *Orthosiphon stamineus* Methanol Extracts in Rats. *Journal of Ethnopharmacology*, 118(3), 354-360.
- Arbuckle, M. I., Kane, S., Porter, L. M., Seatter, M. J. and Gould, G. W. (1996). Structure-Function Analysis of Liver-Type (GLUT2) and Brain-Type (GLUT3) Glucose Transporters: Expression of Chimeric Transporters in *Xenopus* Oocytes Suggests an Important Role for Putative Transmembrane Helix 7 in Determining Substrate Selectivity. *Biochemistry*, 35(51), 16519-16527.
- Armutcu, F., Ataymen, M., Atmaca, H. and Gurel, A. (2008). Oxidative Stress Markers, C-reactive Protein and Heat Shock Protein 70 Levels in Subjects with Metabolic Syndrome. *Clinical Chemistry and Laboratory Medicine*, 46(6), 785-790.
- Assimakopoulos-Jeannet, F. (2004). Fat Storage in Pancreas and in insulin-Sensitive Tissues in Pathogenesis of Type 2 Diabetes. *International Journal of Obesity*, 28, S53-S57.
- Arthur, H. (1954). A Phytochemical Survey of Some Plants of North Borneo. *Journal of Pharmacy and Pharmacology*, 6(1), 66-72.
- Awad, R. (2008). *Ethnopharmacology of Medicinal Plants Used in Anxiety and Epilepsy: Effects on Gamma-Aminobutyric Acid (GABA) Metabolism*. NR50717 Ph.D., University of Ottawa, Canada.
- Ayodhya, S., Kusum, S. and Anjali, S. (2010). Hypoglycaemic Activity of Different Extracts of Various Herbal Plants, *IJRAP*, 1 (1) 212-224
- Ayyanar, M. and Ignacimuthu, S. (2011). Ethnobotanical Survey of Medicinal Plants Commonly Used by Kani Tribals in Tirunelveli Hills of Western Ghats, India. *Journal of Ethnopharmacology*, 134(3), 851-864.
- Aziz, R. A., Sarmidi, M. R., Kumaresan, S., Taher, Z. M. and Foo, D. (2003). Phytochemical Processing: The Next Emerging Field in Chemical Engineering—Aspects and Opportunities. *Jurnal Kejuruteraan Kimia Malaysia*, 3, 45-60.
- Bailey, C. (1988). Metformin Revisited: Its Actions and Indications for Use. *Diabetic medicine*, 5(4), 315-320.
- Bailey, C. J. and Day, C. (1989). Traditional Plant Medicines as Treatments for Diabetes. *Diabetes Care*, 12(8), 553-564.

- Barutta, F., Pinach, S., Giunti, S., Vittone, F., Forbes, J. M., Chiarle, R., ... & Gruden, G. (2008). Heat Shock Protein Expression in Diabetic Nephropathy. *American Journal of Physiology-Renal Physiology*, 295(6), 1817-1824.
- Baynes, J. W. (1991). Role of Oxidative Stress in Development of Complications in Diabetes. *Diabetes*, 40(4), 405-412.
- Bedoya, F., Wilson, J., Ghosh, A., Finegold, D. and Matschinsky, F. (1986). The Glucokinase Glucose Sensor in Human Pancreatic Islet Tissue. *Diabetes*, 35(1), 61-67.
- Bhikha, R. and Glynn, J. (2013). Diabetes: What, Why and How Tibb Can Help. Tibb Institute, Ascience of Medicine, 2-7.
- Bhuvaneswari, P., & Krishnakumari, S. (2012). Nephroprotective Effects of Ethanolic Extract of Sesamum Indicum Seeds (Linn.) in Streptozotocin Induced Diabetic Male Albino Rats. *International Journal of Green Pharmacy*, 6(4), 330.
- Boizel, R., Benhamou, P. Y., Lardy, B., Laporte, F., Foulon, T. and Halimi, S. (2000). Ratio of Triglycerides to HDL Cholesterol is an Indicator of LDL Particle Size in Patients with Type 2 Diabetes and Normal HDL Cholesterol Levels. *Diabetes Care*, 23(11), 1679-1685.
- Bonnefont-Rousselot, D. (2002). Glucose and Reactive Oxygen Species. *Current Opinion in Clinical Nutrition & Metabolic Care*, 5(5), 561-568.
- Bonnefont-Rousselot, D. (2004). The Role of Antioxidant Micronutrients in the Prevention of Diabetic Complications. *Treatments in Endocrinology*, 3(1), 41-52.
- Bradford, MM. (1976). A Rapid and Sensitive for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry* 72: 248-254..
- Brevoort, P. (1998). The Booming US Botanical Market; A New Overview. *Herbalgram*. 1998;44:33-46.
- Brownlee, M. (2001). Biochemistry and Molecular Cell Biology of Diabetic Complications. *Nature*, 414(6865), 813-820.
- Bruce, C. R., Carey, A. L., Hawley, J. A. and Febbraio, M. A. (2003). Intramuscular Heat Shock Protein 72 and Heme Oxygenase-1 mRNA are Reduced in Patients With Type 2 Diabetes Evidence That Insulin Resistance Is



- Associated With a Disturbed Antioxidant Defense Mechanism. *Diabetes*, 52(9), 2338-2345.
- Bublin, M., Radauer, C., Wilson, I. B. H., Kraft, D., Scheiner, O., Breiteneder, H., & Hoffmann-sommergruber, K. (2003). from allergic patients. *The FASEB Journal*, 4.
- Bulbul, I. J., Ullah, M. O., Rahman, M. A., Rahman, K. A., Paul, A. K. and Choudhuri, M. (2009). Effect of “Garbha Cintamani Rasa”, an Ayurvedic Formulation on Lipid Profile, Liver Function and Kidney Function Parameters of Rat Plasma after Chronic Administration. *European Journal of Scientific Research*, 32(1), 25-32.
- Burkill, I. H. (1935). *Dictionnary of the Economic Products of the Malay Peninsula*: Ministry of Agriculture (Malaysia). Crown Agents for the Colonies. London. 839.
- Bourqui, R. M. (2012). Carnitine: Analytical and Physiological Aspects. PhD Thesis, University of Basel, Germany.
- Bwin, M., Bwin, D. M. and Gwan, S. (1967). *Plants with Reputed Hypoglycemic Action: Health and Myanmar Traditional Medicine*,” Burma Medical Research Institute, Ministry of Health, Yangon, pp. 126—128.
- Cai, Y., Luo, Q., Sun, M. and Corke, H. (2004). Antioxidant Activity and Phenolic Compounds of 112 Traditional Chinese Medicinal Plants Associated with Anticancer. *Life sciences*, 74(17), 2157-2184.
- Cai, Y., Sun, M. and Corke, H. (2003). Antioxidant Activity of Betalains from Plants of the Amaranthaceae. *Journal of Agricultural and Food Chemistry*, 51(8), 2288-2294.
- Carrick, J., Chan, K. and Cheung, H. (1968). A New Phytochemical Survey of Malaya-Chemical Screening. *Chemical & Pharmaceutical Bulletin*, 16(12), 2436-2441.
- Cauthen, C. A., Tong, M., Wu, Y., Jain, A., & Tang, W. H. (2011). Prognostic Importance of Serial Evaluation of Blood Urea Nitrogen in Ambulatory Patients with Chronic Systolic Heart Failure and Preserved Renal Function. *Journal of Cardiac Failure*, 17(8), S86.

- Ceriello, A. (2003). New Insights on Oxidative Stress and Diabetic Complications May Lead to a “Causal” Antioxidant Therapy. *Diabetes Care*, 26(5), 1589-1596.
- Chakravarthy, U., Hayes, R. G., Stitt, A. W., McAuley, E. and Archer, D. B. (1998). Constitutive Nitric Oxide Synthase Expression in Retinal Vascular Endothelial Cells is Suppressed by High Glucose and Advanced Glycation End Products. *Diabetes*, 47(6), 945-952.
- Chan, K. and Teo, L. (1969). A New Phytochemical Survey of Malaya. II. Chemical Screening. *Chemical & Pharmaceutical Bulletin*, 17(6), 1284.
- Chan, K. and Teo, L. (1972). A New Phytochemical Survey of Malaya. III. Chemical Screening. *Chemical & Pharmaceutical Bulletin*, 20(7), 1582-1584.
- Chanwitheesuk, A., Teerawutgulrag, A. and Rakariyatham, N. (2005). Screening of Antioxidant Activity and Antioxidant Compounds of Some Edible Plants of Thailand. *Food Chemistry*, 92(3), 491-497.
- Che Nor Din, N. (2010). *Use of Complementary and Alternative Medicine among Cancer Patients at Local Hospital in Malaysia*. Universiti Teknologi Mara, Malaysia.
- Chen, H. Y., Lin, Y. C. and Hsieh, C. L. (2007). Evaluation of Antioxidant Activity of Aqueous Extract of Some Selected Nutraceutical Herbs. *Food Chemistry*, 104(4), 1418-1424.
- Chen, H. Y. and Yen, G. C. (2007). Antioxidant Activity and Free Radical-Scavenging Capacity of Extracts from Guava (*Psidium guajava* L.) Leaves. *Food Chemistry*, 101(2), 686-694.
- Chen H, Wu Y, Zhang Y . (2006) Hsp70 inhibits lipopolysaccharide-induced NF-kappaB activation by interacting with TRAF6 and inhibiting its ubiquitination. *FEBS Lett* 580:3145–3152
- Cheng, X. (2004). *The Role of Inducible Nitric Oxide Synthase (iNOS) on Cardiovascular Function in Rats with Streptozotocin-induced Diabetes*. NQ90160 Ph.D., The University of British Columbia, Canada.
- Chin, J.H, S. Ismail and A.H. Hussin. (2008). Induction of Hepatic Glutathione-S-Transferase Activity by Orthosiphon Stamineus, Benth in Stz-Induced Diabetic Rats , *Malaysian Journal of Pharmaceutical Sciences*, Vol. 6, No. 1, 59–68.

- Chung, J., Nguyen, A.-K., Henstridge, D. C., Holmes, A. G., Chan, M. S., Mesa, J. L. (2008). HSP72 Protects Against Obesity-induced Insulin Resistance. *Proceedings of the National Academy of Sciences*, 105(5), 1739-1744.
- Cotelle, N., Bernier, J.-L., Catteau, J.-P., Pommery, J., Wallet, J.-C. and Gaydou, E. M. (1996). Antioxidant Properties of Hydroxy-Flavones. *Free Radical Biology and Medicine*, 20(1), 35-43.
- Craven PA, DeRubertis FR, Melhem M.( 1997). Nitric oxide in diabetic nephropathy. *Kidney Int.* 52:Suppl 60. S46–S53.
- Daily, G. C. (1997). *Nature's Services: Societal Dependence on Natural Ecosystems*: Island Press, Washington, D.C.
- Daily, G. C., Alexander, S., Ehrlich, P. R., Goulder, L., Lubchenco, J., Matson, P. A. (1997). *Ecosystem Services: Benefits Supplied to Human Societies by Natural Ecosystems* (Vol. 2): Ecological Society of America Washington (DC).
- Dawson, K. G., Gomes, D., Gerstein, H., Blanchard, J. F. and Kahler, K. H. (2002). The Economic Cost of Diabetes in Canada, 1998. *Diabetes Care*, 25(8), 1303-1307.
- De Bock, M., Derraik, J. G. and Cutfield, W. S. (2012). Polyphenols and Glucose Homeostasis in Humans. *Journal of the Academy of Nutrition and Dietetics*, 112(6), 808-815.
- De Matos, L. L., Trufelli, D. C., de Matos, M. G. L. and da Silva Pinhal, M. A. (2010). Immunohistochemistry as an Important Tool in Biomarkers Detection and Clinical Practice. *Biomarker Insights*, 5, 9.
- Dobbins, R. L., Chester, M. W., Stevenson, B. E., Daniels, M. B., Stein, D. T. and McGarry, J. D. (1998). A Fatty Acid-Dependent Step is Critically Important for both Glucose-and Non-G -Stimulated Insulin Secretion. *Journal of Clinical Investigation*, 101(11), 2370.
- Dool, C. J. (2009). Pharmacologic Inhibition of Insulin Receptor Tyrosine Kinase Activity Has Antineoplastic Effects Similar to Alloxan-induced Insulin Deficiency with Less Acute Metabolic Toxicity.
- Donga, J., Surani, V., Sailor, G., Chauhan, S. and Seth, A. (2011). A Systematic Review on Natural Medicine used for Therapy of Diabetes Mellitus of Some Indian Medicinal Plants. *International Journal of Pharmaceutical Sciences*, 2(1), 36-72.

- Douglas, B. and Kiang, A. (1957). A Phytochemical Survey of Malaya. *Malayan Pharmaceutical Journal*, 6, 1-16.
- Du, Y., Smith, M. A., Miller, C. M. and Kern, T. (2002). Diabetes-Induced Nitrate Stress in the Retina, and Correction by Aminoguanidine. *Journal of Neurochemistry*, 80(5), 771-779.
- Duh, P.D. (1998). Antioxidant Activity of Burdock (*Arctium Lappa* Linne): Its Scavenging Effect on Free-Radical and Active Oxygen. *Journal of the American Oil Chemists' Society*, 75(4), 455-461.
- Dunn, J. S., Duffy, E., Gilmour, M., Kirkpatrick, J. and McLetchie, N. (1944). Further Observations on the Effects of Alloxan on the Pancreatic Islets. *The Journal of physiology*, 103(2), 233-243.
- Duraisamy, Y., Gaffney, J., Slevin, M., Smith, C. A., Williamson, K. and Ahmed, N. (2003). Aminosalicic Acid Reduces the Antiproliferative Effect of Hyperglycaemia, Advanced Glycation Endproducts And Glycated Basic Fibroblast Growth Factor In Cultured Bovine Aortic Endothelial Cells: Comparison With Aminoguanidine *Vascular Biochemistry* (pp. 143-153): Springer.
- Ebaid, H., Salem, A., Sayed, A. and Metwalli, A. (2011). Whey Protein Enhances Normal Inflammatory Responses during Cutaneous Wound Healing in Diabetic Rats. *Lipids Health Dis*, 10(14), 235.
- Elavarasi, S., Saravanan, K. and Renuka, C. (2013). A Systematic Review on Medicinal Plants Used to Treat Diabetes Mellitus. 3(3), 983-992.
- Erekat, N., Al Khatib, A. and Al-Jarrah, M. (2014). Endurance Exercise Training Attenuates the up Regulation of iNOS in the Skeletal Muscles of Chronic/Progressive Mouse Model of Parkinson's Disease. *Journal of Neurology Research*, 3(3-4), 108-113.
- Evans, J. L., Goldfine, I. D., Maddux, B. A. and Grodsky, G. M. (2002). Oxidative Stress and Stress-Activated Signaling Pathways: A Unifying Hypothesis of Type 2 Diabetes. *Endocrine Reviews*, 23(5), 599-622.
- Evans, J. L., Goldfine, I. D., Maddux, B. A. and Grodsky, G. M. (2003). Are Oxidative Stress- Activated Signaling Pathways Mediators of Insulin Resistance and  $\beta$ -Cell Dysfunction? *Diabetes*, 52(1), 1-8.

- Farnsworth, N., Blowster, R., Darmratoski, D., Meer, W. and Cammarato, L. (1967). Studies on Catharanthus alkaloids IV Evaluation by means of TLC and Ceric Ammonium Sulphate Spray Reagent. *Lloydia*, 27, 302-314.
- Farnsworth, N. R. and Bingel, A. (1977). Problems and Prospects of Discovering New Drugs from Higher Plants by Pharmacological Screening *New Natural Products and Plant Drugs with Pharmacological, Biological or Therapeutical Activity* (pp. 1-22): Springer.
- Federici, M., Hribal, M., Perego, L., Ranalli, M., Caradonna, Z., Perego, C. (2001). High Glucose Causes Apoptosis in Cultured Human Pancreatic Islets of Langerhans A Potential Role for Regulation of Specific Bcl Family Genes Toward an Apoptotic Cell Death Program. *Diabetes*, 50(6), 1290-1301.
- Felig P, Bergaman M. The endocrine pancreas: Diabetes mellitus.( 1995) In: Felig P, Baxter JD, Frohman LA, editors. *Endocrinology and Metabolism*. New York: M`cGraw-Hill;. p. 1107-250.
- Fiedeler, U. and Nentwich, M. Nanoparticles. (2012) Free Radicals and Oxidative Stress. *NanoTrust Dossiers*, 12, 1-3.
- Firuzi, O., Lacanna, A., Petrucci, R., Marrosu, G. and Saso, L. (2005). Evaluation of the Antioxidant Activity Of Flavonoids by “Ferric Reducing Antioxidant Power” Assay and Cyclic Voltammetry. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1721(1), 174-184.
- Fortes, M. B., & Whitham, M. (2009). No Endogenous Circadian Rhythm in Resting Plasma Hsp72 Concentration in Humans. *Cell Stress and Chaperones*, 14(3), 273-280.
- Fossati, P. and and Prencipe, L. (1982). Estimation of the Concentration of Triglyceride in Plasma and Liver. *Clinical Chemistry* 28: 2077- 2081.
- Fraser, M.-H. (2006). *Ethnobotanical Investigation of Plants Used for the Treatment of Type 2 Diabetes by Two Cree Communities in Quebec: Quantitative Comparisons and Antioxidant Evaluation*. MR24669 M.Sc., McGill University ,Canada.
- Fridlyand, L. E. and Philipson, L. H. (2004). Does the Glucose-Dependent Insulin Secretion Mechanism Itself Cause Oxidative Stress in Pancreatic  $\beta$ -cells? *Diabetes*, 53(8), 1942-1948.

- Ganugapati, J., Baldwa, A. and Lalani, S. (2012). Molecular Docking Studies of Banana Flower Flavonoids As Insulin Receptor Tyrosine Kinase Activators As a Cure for Diabetes Mellitus. *Bioinformation*, 8(5), 216.
- Gavin, J., Alberti, K., Davidson, M., DeFronzo, R., Drash, A., Gabbe, S. (1997). Report of the Expert Committee on the Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care*, 20(7), 1183-1197.
- Gazzaneo, L. R. S., De Lucena, R. F. P. and De Albuquerque, U. P. (2005). Journal of Ethnobiology and Ethnomedicine. *Journal of Ethnobiology and Ethnomedicine*, 1, 9.
- Ghorbani, A., Naghibi, F. and Mosaddegh, M. (2006). Ethnobotany, Ethnopharmacology and drug discovery. *Iranian J. Pharm. Sci*, 2, 109-118.
- Ghous, T., Akhtar, K., Nasim, F. U. H., & Choudhry, M. A. (2010). Screening of selected medicinal plants for urease inhibitory activity. *Biology & Medicine*, 2(4).
- Ginsberg, H. N., Elam, M. B., Lovato, L. C., Crouse 3rd, J., Leiter, L. A., Linz, P. (2010). Effects of combination Lipid Therapy In Type 2 Diabetes Mellitus. *The New England journal of medicine*, 362(17), 1563-1574.
- Goh, T. T., Mason, T. M., Gupta, N., So, A., Lam, T. K., Lam, L. (2007). Lipid-Induced  $\beta$ -cell Dysfunction In Vivo in Models of Progressive  $\beta$ -cell Failure. *American Journal of Physiology-Endocrinology And Metabolism*, 292(2), E549-E560.
- Goodenow, T. J. and Malarkey, W. B. (1977). Leukocytosis and Artifactual Hypoglycemia. *JAMA: The Journal of the American Medical Association*, 237(18), 1961-1962.
- Griendling, K. K. and Fitz Gerald, G. A. (2003). Oxidative Stress and Cardiovascular Injury Part I: Basic Mechanisms And In Vivo Monitoring of ROS. *Circulation*, 108(16), 1912-1916.
- Group, D. R. (1994). Effect of intensive Diabetes Treatment on the Development and Progression of Long-Term Complications in Adolescents with Insulin-Dependent Diabetes Mellitus: Diabetes Control and Complications Trial. *J pediatr*, 125(2), 177-188.
- Group, N. D. D. (1979). Classification and Diagnosis of Diabetes Mellitus and Other Categories of Glucose Intolerance. *Diabetes*, 28(12), 1039-1057.

- Grundy, S. M., Benjamin, I. J., Burke, G. L., Chait, A., Eckel, R. H., Howard, B. V. (1999). Diabetes and Cardiovascular Disease: A Statement for Healthcare Professionals from the American Heart Association. *Circulation*, 100(10), 1134-1146.
- Gugliucci, A., Hermo, R., Monroy, C., Numaguchi, M., & Kimura, S. (2005). Ischemia-modified Albumin Levels in Cord Blood: A Case-Control Study in Uncomplicated and Complicated Deliveries. *Clinica chimica acta*, 362(1), 155-160.
- Gulliford, M. C. (1994). News from Overseas: Health and Health Care in the English-Speaking Caribbean: A British Public Health Physician's View of the Caribbean. *Journal of Public Health*, 16(3), 263-269.
- Gullo, C. A. and Teoh, G. (2004). Heat Shock Proteins: to Present or Not, That is the Question. *Immunology letters*, 94(1), 1-10.
- Gupte, A. A. (2009). *Heat Shock Proteins: Novel Therapeutic Targets Against Insulin Resistance and Type 2 Diabetes*. 3355995 Ph.D., University of Kansas, United state.
- Halliwell, B. (1994). Free Radicals, Antioxidants, and Human Disease: Curiosity, Cause, orC? *The Lancet*, 344(8924), 721-724.
- Harmon, J. S., Gleason, C. E., Tanaka, Y., Poitout, V. and Robertson, R. P. (2001). Antecedent Hyperglycemia, Not Hyperlipidemia, is Associated with Increased Islet Triacylglycerol Content and Decreased Insulin Gene mRNA Level in Zucker Diabetic Fatty Rats. *Diabetes*, 50(11), 2481-2486.
- Heilbronn, L., Smith, S. and Ravussin, E. (2004). Failure of Fat Cell Proliferation, Mitochondrial Function and Fat Oxidation Results in Ectopic Fat Storage, Insulin Resistance and Type II Diabetes Mellitus. *International Journal of Obesity*, 28, S12-S21.
- Heinrich, M., Ankli, A., Frei, B., Weimann, C. and Sticher, O. (1998). Medicinal in Mexico: Healers' Consensus and Cultural Importance. *Social Science & Medicine*, 47(11), 1859-1871.
- Henry, R. J., Cannon, D. C. and Winkelman, J. W. (1974). *Clinical Chemistry: Principles and Technics* (Vol. 525): Harper & Row Hagerstown, New York, Evanston, San Francisco, London.
- Hickman, I. J. and Macdonald, G. A. (2007). Impact of Diabetes on the Severity of Liver Disease. *The American Journal of Medicine*, 120(10), 829-834.

- Hii, C. and Howell, S. (1985). Effects of Flavonoids on Insulin Secretion and  $^{45}\text{Ca}^{2+}$  Handling in Rat Islets of Langerhans. *Journal of Endocrinology*, 107(1), 1-8.
- Hoareau, L., & DaSilva, E. (1999). Medicinal plants: a re-emerging health aid. *Electronic Journal Of Biotechnology*, 2(2).
- Huang, D. Ou, B.; Prior, R.L.( 2005). The Chemistry behind Antioxidant Capacity Assays. *J. Agric. Food Chem.* 53, 1841–1856.
- Hudaib, M., Mohammad, M., Bustanji, Y., Tayyem, R., Yousef, M., Abuirjeie, M. (2008). Ethnopharmacological Survey of Medicinal Plants in Jordan, Mujib Nature Reserve and Surrounding Area. *Journal of Ethnopharmacology*, 120(1), 63-71.
- Ibrahim Jantan, (2004). Medicinal plant research in Malaysia: scientific interests and advances. *Jurnal Sains Kesihatan Malaysia*, 2 (2). pp. 27-46. ISSN 1675-8161.
- Igbinaduwa, P., Igbinaduwa, B., and Oforofuo, I. (2008). Lipid Concentration And The Extent Of Their Peroxidation In Nigerian Hypertensives. *Internet Journal of Health*, 8(1).
- Irshaid, F., Jaran, A., Dilmi, F., Tarawneh, K., Hadeth, R. and Al-Khatib, A. (2010). Prevalence of Epstein Barr Virus Latent Membrane Protein-1 in Jordanian Patients with Hodgkin's Lymphoma and Non-Hodgkin's Lymphoma. *J Biol Sci*, 10, 507-513.
- Irshaid, F. and Mansi, K. (2009). The Effects of Methanol Extract Derived From *Urtica Pilulifera* Leaves on Some Hematological and Biochemical Parameters Of Diabetic Rats. *Res. J. Biol. Sci*, 4, 675-681.
- Jacob, R. A. (1995). The Integrated Antioxidant System. *Nutrition Research*, 15(5), 755-766.
- Jacqueminet, S., Briaud, I., Rouault, C., Reach, G. and Poitout, V. (2000). Inhibition of Insulin Gene Expression by Long-Term Exposure of Pancreatic  $\beta$  Cells to Palmitate is Dependent on the Presence of a Stimulatory Glucose Concentration. *Metabolism*, 49(4), 532-536.
- Jacquier-Sarlin, M., Fuller, K., Dinh-Xuan, A., Richard, M.-J. and Polla, B. (1994). Protective Effects of Hsp70 in Inflammation. *Experientia*, 50(11-12), 1031-1038.



- Jamal, J. A. (2006). Malay Traditional Medicine. *Tech Monitor (Special Feature: Traditional Medicine: S & T Advancement)*, pp, 37-49.
- Jantan, I. (2004). Medicinal Plant Research in Malaysia: Scientific Interests and Advances. *Jurnal Sains Kesihatan Malaysia*, 2(2), 27-46.
- Jayaprakasam, B., Vareed, S. K., Olson, L. K. and Nair, M. G. (2005). Insulin Secretion by Bioactive Anthocyanins and Anthocyanidins Present in Fruits. *Journal of Agricultural and Food Chemistry*, 53(1), 28-31.
- Joharchi, K., & Jorjani, M. (2007). The Role of Nitric Oxide in Diabetes-Induced Changes of Morphine Tolerance in Rats. *European Journal of Pharmacology*, 570(1), 66-71.
- Kahn, S. E., Hull, R. L. and Utzschneider, K. M. (2006). Mechanisms Linking Obesity to Insulin Resistance and Type 2 Diabetes. *Nature*, 444(7121), 840-846.
- Kaya, G. (2006). Discussion of P&P Model Used For Estimating Option Value of Forest Ecosystems as Medicinal Plant Resources. *Journal of the Bartın Faculty of Forestry*, 23-32.
- Keen, R., Deacon, A., Delves, H., Moreton, J. and Frost, P. (1994). Indian Herbal Remedies for Diabetes as a Cause of Lead Poisoning. *Postgraduate medical journal*, 70(820), 113-114.
- Keenan, T., Joe, P., Wilson, J., Collier, C., Golding, B., Burgess, D. (2003). The Sydney 2000 World Weather Research Programme Forecast Demonstration Project: Overview and Current Status. *Bulletin of the American Meteorological Society*, 84(8), 1041-1054. doi: 10.1175/bams-84-8-1041
- Kemertelidze, E., Sagareishvili, T., Syrov, V., Khushbaktova, Z., Tsutskiridze, L. and Kurashvili, R. (2012). Saturin-Effective Vegetative Remedy in Treatment of Type 2 Diabetes Mellitus. *Georgian Medical News*, (203), 47.
- Kennedy, L. and Baynes, J. (1984). Non-enzymatic glycosylation and the chronic complications of diabetes: an overview. *Diabetologia*, 26(2), 93-98.
- Khatun, M. A., Harun-Or-Rashid, M. and Rahmatullah, M. (2011). Scientific Validation of Eight Medicinal Plants Used in Traditional Medicinal Systems of Malaysia: A Review. *American-Eurasian Journal of Sustainable Agriculture*, 5(1), 67-75.

- Khatun, M. H., Nesa, M. L., Islam, R., Ripa, F. A., & Kadir, S. (2014). Antidiabetic and antidiarrheal effects of the methanolic extract of *Phyllanthus reticulatus* leaves in mice. *Asian Pacific Journal of Reproduction*, 3(2), 121-127.
- Kiang, A., Douglas, B. and Morsingh, F.(1961). A Phytochemical Survey of Malaya: Part II. Alkaloids. *Journal of Pharmacy and Pharmacology*, 13(1), 98-104.
- Kim, W., & Egan, J. M. (2008). The Role of Incretins in Glucose Homeostasis and Diabetes Treatment. *Pharmacological Reviews*, 60(4), 470-512.
- King, H., Aubert, R. E. and Herman, W. H. (1998). Global Burden of Diabetes, 1995–2025: Prevalence, Numerical Estimates, and Projections. *Diabetes Care*, 21(9), 1414-1431.
- King, A. J. (2012). The Use of Animal Models in Diabetes Research. *British Journal of Pharmacology*, 166(3), 877-894.
- Knekt, P., Kumpulainen, J., Järvinen, R., Rissanen, H., Heliövaara, M., Reunanen, A., Hakulinen T, Aromaa A. (2002). Flavonoid intake and Risk of Chronic Diseases. *The American Journal of Clinical Nutrition*, 76(3), 560-568.
- Kobayashi, K., Baba, E., Fushiya, S., Takano, F., Batkhuu, J., Dash, T. (2003). Screening of Mongolian Plants for Influence on Amylase Activity in Mouse Plasma and Gastrointestinal Tube. *Biological And Pharmaceutical Bulletin*, 26(7), 1045-1048.
- Kodama, T., Arimoto, N., Barger, A. J. and Aragon-Salamanca, A. (1998). Evolution of the Colour-Magnitude Relation of Early-Type Galaxies in Distant Clusters. *Arxiv Preprint Astro-ph/9802245*.
- Kohen, R. and Nyska, A. (2002). Invited Review: Oxidation of Biological Systems: Oxidative Stress Phenomena, Antioxidants, Redox Reactions, and Methods For Their Quantification. *Toxicologic Pathology*, 30(6), 620-650.
- Koh, H., Tasaka, S., Hasegawa, N., Yamada, W., Shimizu, M., Nakamura, M. & Ishizaka, A. (2007). Protective Role of Vascular Endothelial Growth Factor in Endotoxin-Induced Acute Lung Injury in Mice. *Respir Res*, 8(1), 60.
- Krippner, S. (2003). Models of Ethnomedicinal Healing. Proceedings of the 2003 *Ethnomedicine Conferences, Munich, Germany. April, 26-27*.
- Kulip, J. (2003). An Ethnobotanical Survey of Medicinal and Other Useful Plants of Muruts in Sabah, Malaysia. *Telopea*, 10(1), 81-98.

- Kulip, J., Lam, N. F., Manshoor, N., Julius, A., Said, I. M., Gisil, J., Julianah, A.J, Welly, F.Tukin. (2010). Medicinal Plants in Maliau Basin, Sabah, Malaysia. *Journal Of Tropical Biology And Conservation*, 6, 21-33.
- Kumar, G. L., & Rudbeck, L. (Eds.). (2009). Education Guide: Immunohistochemical Staining Methods: Pathology. Dako North America.
- Kurucz, I., Morva, Á., Vaag, A., Eriksson, K.-F., Huang, X., Groop, L., Koranyi L. (2002). Decreased Expression of Heat Shock Protein 72 in Skeletal Muscle of Patients with Type 2 Diabetes Correlates with Insulin Resistance. *Diabetes*, 51(4), 1102-1109.
- Langseth, L. (1993). From the Editor: Antioxidants and Diseases of the Brain. *Antioxidant Vitamins Newsletter*, 4, 3.
- Larkins, N. and Wynn, S. (2004). Pharmacognosy: Phytomedicines and Their Mechanisms. *Veterinary Clinics of North America: Small Animal Practice*, 34(1), 291-327.
- Larson, R. A. (1988). The Antioxidants of Higher Plants. *Phytochemistry*, 27(4), 969-978.
- Lee, H. W., Hakim, P., Rabu, A., & Sani, H. A. (2012). Antidiabetic Effect of Gynura Procumbens Leaves Extracts Involve Modulation of Hepatic Carbohydrate Metabolism in Streptozotocin-Induced Diabetic Rats. *J Med Plants Res*, 6(5), 796-812.
- Lee, S., Xiao, C. and Pei, S. (2008). Ethnobotanical Survey of Medicinal Plants at Periodic Markets of Honghe Prefecture in Yunnan Province, SW China. *Journal of Ethnopharmacology*, 117(2), 362-377.
- Leiter LA, Barr A, Bélanger A, Lubin S, Ross SA, Tildesley HD, Fontaine N; Diabetes Screening in Canada (DIASCAN) Study. (2001). Diabetes Screening in Canada (DIASCAN) Study: Prevalence of Undiagnosed Diabetes and Glucose Intolerance in Family Physician Offices. *Diabetes Care*, 24, 1038-1043.
- Leonardi, O., Mints, G. and Hussain, M. A. (2003). Beta-cell Apoptosis in the Pathogenesis of Human Type 2 Diabetes Mellitus. *European Journal of Endocrinology*, 149(2), 99-102.
- Lenzen, S. (2008). The Mechanisms of Alloxan-and Streptozotocin-Induced Diabetes. *Diabetologia*, 51(2), 216-226.

- Lenzen, S., Tiedge, M., Jörns, A. and Munday, R. (1996). Alloxan Derivatives as a Tool for the Elucidation of the Mechanism of the Diabetogenic Action of Alloxan *Lessons from Animal Diabetes VI* (pp. 113-122): Springer.
- Li, W., Zhong, X., Zhang, L., Wang, Y., & Wang, T. (2012). Heat Shock Protein 70 Expression is Increased in the Liver of Neonatal Intrauterine Growth Retardation Piglets. *Asian-Australasian Journal of Animal Sciences*, 25(8), 1096.
- Lowe, H., Payne-Jackson, A., Beckstrom-Sternberg, S. M. and Duke, J. A. (2001). *Jamaica's Ethnomedicine: Its Potential in the Healthcare System*: Pelican Publishers.
- Lukačinová, A., Mojlić, J., Beňačka, R., Rácz, O. and Niatiar, F. (2008). Structure-Activity Relationships of Preventive Effects of Flavonoids in Alloxan-Induced Diabetes Mellitus in Rats. *Journal of Animal and Feed Sciences*, 474, 46.
- Lupi, R., Dotta, F., Marselli, L., Del Guerra, S., Masini, M., Santangelo, C., Patané, G., Boggi, U., Piro, S., Anello, M., Bergamini, E., Mosca, F., Di Mario, U., Del Prato, S., Marchetti, P. (2002). Prolonged Exposure to Free Fatty Acids Has Cytostatic and Pro-Apoptotic Effects on Human Pancreatic Islets: Evidence that  $\beta$ -Cell Death Is Caspase Mediated, Partially Dependent on Ceramide Pathway, and Bcl-2 Regulated. *Diabetes*, 51(5), 1437-1442.
- Lyon, H., Andersen, A. P., Hasselager, E., Høyer, P. E., Møller, M., Prentø, P., & Van Deurs, B. (1991). *Theory and Strategy in Histochemistry*, Berlin: Springer Verlag.
- Malviya, N., Jain, S. and Malviya, S. (2010). Antidiabetic Potential of Medicinal Plants. *Acta Pol Pharm*, 67(2), 113-118.
- Manna, P., Das, J., Ghosh, J. and Sil, P. C. (2010). Contribution of Type 1 Diabetes to Rat Liver Dysfunction and Cellular Damage Via Activation of NOS, PARP, I $\kappa$ B $\alpha$ /NF- $\kappa$ B, MAPKs, and Mitochondria-Dependent Pathways: Prophylactic Role of Arjunolic Acid. *Free Radical Biology and Medicine*, 48(11), 1465-1484.
- Manosroi, A. Saraphanchotiwiththaya, A.; Manosroi, J. (2003). Immunomodulatory Activities of *Clausena Excavata* Burm. f. Wood Extracts. *J. Ethnopharmacol.* 89, 155–160.

- Mansi, K., Amneh, M. and Nasr, H. (2007). The Hypolipidemic Effects of *Artemisia Sieberi* (A. *Herba-Alba*) in Alloxan Induced Diabetic Rats. *Inter. J. Pharm*, 3, 487-491.
- Mansi, K., and Lahham, J. (2008). Effects of *Artemisia Sieberi* Besser (A. *herba-alba*) on Heart Rate and Some Hematological Values in Normal and Alloxan-Induced Diabetic Rats. *J. Basic Appl. Sci*, 4(2), 57-62.
- Marinelli, J. (2005). Plant: The Ultimate Visual Reference to Plants and Flowers of the World. *New York, NY: DK*.
- Marletta, M. A. 1993. Nitric Oxide Synthase Structure and Mechanism, *J Biol Chem* 268:12231-12234.
- Marouf, S. A. H. and Hasan, B. (2013). Flavonoids as Alternatives in Treatment of Type 2 Diabetes Mellitus. *Academia Journal of Medicinal Plants (ISSN 2315-7720)*, 1(2), 031-036.
- Maroyi, A. (2011). An Ethnobotanical Survey of Medicinal Plants Used by the People in Nhema Communal Area, Zimbabwe. *Journal of Ethnopharmacology*, 136(2), 347-354.
- Marra G1, Cotroneo P, Pitocco D, Manto A, Di Leo MA, Ruotolo V, Caputo S, Giardina B, Ghirlanda G, Santini SA. (2002). Early Increase of Oxidative Stress and Reduced Antioxidant Defenses in Patients with Uncomplicated Type 1 Diabetes A Case For Gender Difference. *Diabetes Care*, 25(2), 370-375.
- Martineau LC1, Couture A, Spoor D, Benhaddou-Andaloussi A, Harris C, Meddah B, Leduc C, Burt A, Vuong T, Mai Le P, Prentki M, Bennett SA, Arnason JT, Haddad PS. (2006). Anti-diabetic Properties of the Canadian Lowbush blueberry *Vaccinium Angustifolium* Ait. *Phytomedicine*, 13(9), 612-623.
- Mayer, M. and Bukau, B. (2005). Hsp70 Chaperones: Cellular Functions and Molecular Mechanism. *Cellular and Molecular Life Sciences*, 62(6), 670-684.
- Mayfield, J. (1998). Diagnosis and Classification of Diabetes Mellitus: New Criteria. *American Family Physician*, 58, 1355-1372.
- McCance, D., Hanson, R., Pettitt, D., Bennett, P., Hadden, D. and Knowler, W. (1997). Diagnosing Diabetes Mellitus—Do We Need New Criteria? *Diabetologia*, 40(3), 247-255.

- McCune, L. M. and Johnson, T. (2002). Antioxidant Activity in Medicinal Plants Associated with the Symptoms of Diabetes Mellitus Used by the Indigenous Peoples of the North American Boreal Forest. *Journal of Ethnopharmacology*, 82(2), 197-205.
- McKendry, J. (1988). Direct Costs of Diabetes Care: A Survey in Ottawa, Ontario 1986. *Canadian Journal of Public Health. Revue Canadienne de Sante Publique*, 80(2), 124-128.
- Meglasson, M. D. and Matschinsky, F. M. (1986). Pancreatic Islet Glucose Metabolism and Regulation of Insulin Secretion. *Diabetes/metabolism Reviews*, 2(3-4), 163-214.
- Meltzer S, Leiter L, Daneman D, Gerstein HC, Lau D, Ludwig S, Yale JF, Zinman B, Lillie D. (1998). Clinical Practice Guidelines for the Management of Diabetes in Canada. *Canadian Medical Association Journal*, 159(8), S1-S29.
- Menon, L. G., Kuttan, R. and Kuttan, G. (1999). Anti-metastatic Activity of Curcumin and Catechin. *Cancer Letters*, 141(1), 159-165.
- Mighiu, P. (2012). The Role of Central Glucagon Signaling in the Regulation of Glucose Homeostasis (Doctoral dissertation).
- Miller, R. T. (2001). Technical Immunohistochemistry: Achieving Reliability and Reproducibility of Immunostains. In Society for Applied Immunohistochemistry, Annual Meeting.
- Mohamed, E. A. H., Yam, M. F., Ang, L. F., Mohamed, A. J. and Asmawi, M. Z. (2013). Antidiabetic Properties and Mechanism of Action of Orthosiphon stamineus Benth Bioactive Sub-fraction in Streptozotocin-induced Diabetic Rats. *Journal of Acupuncture and Meridian Studies*, 6(1), 31-40.
- Moller, P. and Loft, S. (2002). Oxidative DNA Damage in Human White Blood Cells in Dietary Antioxidant Intervention Studies. *The American Journal of Clinical Nutrition*, 76(2), 303-310.
- Moran, A., Zhang, H.-J., Olson, L. K., Harmon, J. S., Poitout, V. and Robertson, R. P. (1997). Differentiation of Glucose Toxicity from Beta Cell Exhaustion During the Evolution of Defective Insulin Gene Expression in the Pancreatic Islet Cell Line, HIT-T15. *Journal of Clinical Investigation*, 99(3), 534.
- Morgan, M. (1995). The Significance of Ethnicity for Health Promotion: Patients' Use of Anti-hypertensive Drugs in Inner London. *International Journal of Epidemiology*, 24(Supplement 1), S79-S84.

- Morgan, M. and Watkins, C. (1988). Managing Hypertension: Beliefs and Responses to Medication among Cultural Groups. *Sociology of Health & Illness*, 10(4), 561-578.
- Morino, K., Petersen, K. F. and Shulman, G. I. (2006). Molecular Mechanisms of Insulin Resistance in Humans and Their Potential Links with Mitochondrial Dysfunction. *Diabetes*, 55(Supplement 2), S9-S15.
- Morino K, Neschen S, Bilz S, Sono S, Tsirigotis D, Reznick RM, Moore I, Nagai Y, Samuel V, Sebastian D, White M, Philbrick W, Shulman GI. (2008). Muscle-specific IRS-1 Ser> Ala Transgenic Mice are Protected from Fat-Induced Insulin Resistance in Skeletal Muscle. *Diabetes*, 57(10), 2644-2651.
- Muhammad, B. Y. and Awaisu, A. (2008). The Need for Enhancement of Research, Development, and Commercialization of Natural Medicinal Products in Nigeria: Lessons from the Malaysian Experience. *African Journal of Traditional, Complementary, and Alternative Medicines*, 5(2), 120.
- Muralidhara and Narasimhamurthy, K. (1988). Non-Mutagenicity of Capsaicin in Albino Mice. *Food and Chemical Toxicology*, 26(11-12), 955-958.
- Muranyi, M., He, Q. P., Fong, K. S., & Li, P. A. (2005). Induction of heat shock proteins by hyperglycemic cerebral ischemia. *Molecular brain research*, 139(1), 80-87.
- Najemnikova, E., Rodgers, C. D. and Locke, M. (2007). Altered Heat Stress Response Following Streptozotocin-Induced Diabetes. *Cell Stress & Chaperones*, 12(4), 342.
- Nassl, A. M., Rubio-Aliaga, I., Fenselau, H., Marth, M. K., Kottra, G., & Daniel, H. (2011). Amino Acid Absorption and Homeostasis in Mice Lacking the Intestinal Peptide Transporter PEPT1. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 301(1), G128-G137.
- Nature,(2001). Macmillan Magazines Ltd. VOL 414 | 13 DECEMBER 2001.
- Norhayati M, Noor Hayai MI, Nor Fariza N, Rohani AK, Halimah AS, Sharom MY. (1998). Health Status of Orang Asli (Aborigine) Community in Pos Piah, Sungai Siput, Perak, Malaysia. *Southeast Asian J Trop Med Public Health*, 29(1): 58-61.
- Nna, V. U., Oka, V. O., Aluko, E. O., & Helen, O. T. (2013). Comparative Effects of Aqueous Leaf Extract of *Viscum album* (Mistletoe) and *Aloe vera* gel in the

- Management of Streptozotocin-Induced Diabetes Mellitus. *International Journal of Diabetes Research*, 2(5): 84-89
- Null, Ginsberg, H. N., Elam, M. B., Lovato, L. C., Crouse, J. R., Leiter, L. A. (2010). Effects of Combination Lipid Therapy In Type 2 Diabetes Mellitus. *The New England journal of medicine*, 362(17), 1563-1574.
- Ochei, J. and Kolhatkar, A. (2007). *Medical Laboratory Science Theory and Practice*. 6th reprint: Tata McGraw-Hill New Delhi pp918-923.
- Olaleye, M. T. (2007). Cytotoxicity and Antibacterial Activity of Methanolic Extract of *Hibiscus sabdariffa*. *Journal of Medicinal Plants Research*, 1(1), 9-13.
- Ong, H., Chua, S. and Milow, P. (2011). Ethno-medicinal Plants Used by the Temuan Villagers in Kampung Jeram Kedah, Negeri Sembilan, Malaysia. *Ethno Med*, 5(2):, 95-100
- Ong, H., Norlia, A. and Sorayya, M. (2012). Traditional Knowledge and Usage of Edible Plants Among the Temuan Villagers in Kampung Tering, Kuala Pilah, Negeri Sembilan, Malaysia. *Indian Journal of Traditional Knowledge* 11(1), 161-165.
- Ong, H. C., Ahmad, N. and Milow, P. (2011). Traditional Medicinal Plants Used by the Temuan Villagers in Kampung Tering, Negeri Sembilan, Malaysia. *Ethno Med*, 5(3): 169-173
- Ong, H. C. and Nordiana, M. (1999). Malay Ethno-Medico Botany in Machang, Kelantan, Malaysia. *Fitoterapia*, 70(5), 502-513.
- Ong, H. C. and Norzalina, J. (1999). Malay Herbal Medicine in Gemencheh, Negri Sembilan, Malaysia. *Fitoterapia*, 70(1), 10-14.
- Onozato, M. L., Tojo, A., Goto, A., Fujita, T., & Wilcox, C. S. (2002). Oxidative stress and nitric oxide synthase in rat diabetic nephropathy: effects of ACEI and ARB. *Kidney international*, 61(1), 186-194.
- Onunkwo, G., Akah, P. and Udeala, O. (1996). Studies on *Bridelia ferruginea* leaves (1). Stability and Hypoglycaemic Actions of The Leaf Extract Tablets. *Phytotherapy Research*, 10(5), 418-420.
- Orech, F., Akenga, T., Ochora, J., Friis, H. and Aagaard-Hansen, J. (2005). Potential Toxicity of Some Traditional Leafy Vegetables Consumed in Nyangoma Division, Western Kenya. *Afr J Food Nutr Sc (AJFAND)* 5, 1-13.



- Otaka M., Odashima M., & Watanabe S. (2006). Role of heat shock proteins (molecular chaperones) in intestinal mucosal protection. *Biochem Biophys Res Commun.* 348:1–5.
- Ouslimani, N., Peynet, J., Bonnefont-Rousselot, D., Thérond, P., Legrand, A. and Beaudeux, JL. (2005). Metformin Decreases Intracellular Production Of Reactive Oxygen Species In Aortic Endothelial Cells. *Metabolism*, 54(6), 829-834.
- Oyedemi, S. O., Adewusi, E. A., Aiyegoro, O. A., & Akinpelu, D. A. (2011). Antidiabetic and Haematological Effect of Aqueous Extract of Stem Bark of *Azelaia Africana* (Smith) on Streptozotocin–Induced Diabetic Wistar Rats. *Asian Pacific Journal of Tropical Biomedicine*, 1(5), 353-358.
- Paolisso G, G. A., Tagliamonte MR, Saccomanno F, Salvatore T, Gualdiero P, D’Onofrio MV, Howard BV. (1996). Does free fatty Acid Infusion Impair Insulin Action also Through an Increase in Oxidative Stress? *J Clin Endocrinol Metab* 81:4244–4248.
- Patel, D. K., Prasad, S. K., Kumar, R. and Hemalatha, S. (2012). An Overview on Antidiabetic Medicinal Plants Having Insulin Mimetic Property. *Asian Pacific Journal of Tropical Biomedicine*, 2(4), 320-330.
- Patti, M. E., Butte, A. J., Crunkhorn, S., Cusi, K., Berria, R., Kashyap, S. (2003). Coordinated Reduction of Genes of Oxidative Metabolism in Humans With Insulin Resistance and Diabetes: Potential Role of PGC1 And NRF1. *Proceedings of the National Academy of Sciences*, 100(14), 8466-8471.
- Patton, C. J. and Crouch, S. (1977). Spectrophotometric and Kinetics Investigation of the Berthelot Reaction for the Determination of Ammonia. *Analytical Chemistry*, 49(3), 464-469.
- Patz, A. and Smith, R. (1991). The ETDRS and Diabetes 2000. *Ophthalmology*, 98(5), 739-740.
- Percival, M. (1998). Antioxidants. *Clinical Nutrition Insights, Advanced Nutrition Publications, Inc.* 1998.
- Perez-Jimenez J, Fezeu L and M, T. (2011). Dietary Intake of 337 Polyphenols in French Adults. *The American Journal of Clinical Nutrition*, 93(6), 1220-1228.

- Pham-Huy, L. A., He, H. and Pham-Huy, C. (2008). Free radicals, Antioxidants in Disease and Health. *International Journal of Biomedical Science: IJBS*, 4(2), 89.
- Pick, A., Clark, J., Kubstrup, C., Levisetti, M., Pugh, W., Bonner-Weir, S. (1998). Role of Apoptosis in Failure of Beta-cell Mass Compensation for Insulin Resistance and Beta-Cell Defects in the Male Zucker Diabetic Fatty Rat. *Diabetes*, 47(3), 358-364.
- Pirkkala, L., Nykänen, P., & Sistonen, L. E. A. (2001). Roles of the Heat Shock Transcription Factors in Regulation of the Heat Shock Response and Beyond. *The FASEB Journal*, 15(7), 1118-1131.
- Poitout, V. and Robertson, R. P. (2002). Minireview: Secondary  $\beta$ -cell Failure in Type 2 Diabetes—A Convergence of Glucotoxicity and Lipotoxicity. *Endocrinology*, 143(2), 339-342.
- Prince, P. S., Kamalakkannan, N. and Menon, V. P. (2004). Restoration of Antioxidants by Ethanolic Tinospora Cordifolia in Alloxan-Induced Diabetic Wistar Rats. *Acta Pol Pharm*, 61(4), 283-287.
- Pushparaj, P. N. (2004). Evaluation of the Anti-Diabetic Properties of Averrhoa Bilimbi in Animals with Experimental Diabetes Mellitus. Doctor Philosophy, National University of Singapore.
- Qader, S. W., Abdulla, M. A., Chua, L. S., Najim, N., Zain, M. M. and Hamdan, S. (2011). Antioxidant, Total Phenolic Content and Cytotoxicity Evaluation of Selected Malaysian Plants. *Molecules*, 16(4), 3433-3443.
- Qader, Suhailah Wasman. (2012). Evaluation of Gastroprotective Effect of Selected Malaysian Medicinal Plants on Ethanol-Induced Gastric Ulcer in Rats. PhD Thesis, Universiti Teknologi Malaysia, Skudai.
- Rahimi, R., Nikfar, S., Larijani, B. and Abdollahi, M. (2005). A Review on the Role of Antioxidants in the Management of Diabetes and Its Complications. *Biomedicine & Pharmacotherapy*, 59(7), 365-373.
- Reaven, G. M. (1995). Pathophysiology of Insulin Resistance in Human Disease. *Physiological reviews*, 75(3), 473-486.
- Reddy, A. C. P. and Lokesh, B. R. (1994). Studies on Anti-Inflammatory Activity of Spice Principles and Dietary n-3 Polyunsaturated Fatty Acids on Carrageenan-Induced Inflammation in Rats. *Annals of Nutrition and Metabolism*, 38(6), 349-358.

- Rice-Evans, C. and Miller, N. (1996). Antioxidant Activities of Flavonoids as Bioactive Components of Food. *Biochemical Society Transactions*, 24(3), 790-794.
- Ristow, M., Zarse, K., Oberbach, A., Klötting, N., Birringer, M., Kiehntopf, M. (2009). Antioxidants Prevent Health-Promoting Effects of Physical Exercise in Humans. *Proceedings of the National Academy of Sciences*, 106(21), 8665-8670.
- Ritossa, F. (1996). Discovery of the Heat Shock Response. *Cell Stress & Chaperones*, 1(2), 97.
- Roberson, E. (2008). Medicinal Plants at Risk - Nature's Pharmacy, Our Treasure Chest: Why We Must Conserve Our Natural Heritage. Center for Biological Diversity, Tucson, AZ.
- Robertson, R. P., Harmon, J., Tran, P. O., Tanaka, Y. and Takahashi, H. (2003). Glucose Toxicity in  $\beta$ -cells: Type 2 Diabetes, Good Radicals Gone Bad, and the Glutathione Connection. *Diabetes*, 52(3), 581-587.
- Robertson, R. P., Harmon, J., Tran, P. O. T. and Poitout, V. (2004).  $\beta$ -Cell Glucose Toxicity, Lipotoxicity, and Chronic Oxidative Stress in Type 2 Diabetes. *Diabetes*, 53(suppl 1), S119-S124. doi: 10.2337/diabetes.53.2007.S119
- Robertson, R. P. and Harmon, J. S. (2007). Pancreatic Islet  $\beta$ -cell and Oxidative Stress: The Importance of Glutathione Peroxidase. *FEBS Letters*, 581(19), 3743-3748.
- Rokaya, M. B., Münzbergová, Z. and Timsina, B. (2010). Ethnobotanical Study of Medicinal Plants from the Humla District of Western Nepal. *Journal of Ethnopharmacology*, 130(3), 485-504.
- Rolo, A. P. and Palmeira, C. M. (2006). Diabetes and Mitochondrial Function: Role of Hyperglycemia and Oxidative Stress. *Toxicology and Applied Pharmacology*, 212(2), 167-178.
- Roufail, E., Soulis, T., Boel, E., Cooper, M. and Rees, S. (1998). Depletion of Nitric Oxide Synthase-Containing Neurons in the Diabetic Retina: Reversal by Aminoguanidine. *Diabetologia*, 41(12), 1419-1425.
- Rule, A. D., Larson, T. S., Bergstralh, E. J., Slezak, J. M., Jacobsen, S. J., & Cosio, F. G. (2004). Using Serum Creatinine to Estimate Glomerular Filtration Rate: Accuracy in Good Health and in Chronic Kidney Disease. *Annals of Internal Medicine*, 141(12), 929-937.

- Russ, J. C. (2009). *The Image Processing Cookbook: A Guide to the Processing And Analysis of Scientific, Forensic and Technical Images Using Photoshop - Volume 1*.
- Salmela, P. I., Sotaniemi, E. A., Niemi, M. and Mäentausta, O. (1984). Liver Function Tests in Diabetic Patients. *Diabetes Care*, 7(3), 248-254.
- Saltiel, A. R. and Kahn, C. R. (2001). Insulin Signalling and the Regulation of Glucose and Lipid Metabolism. *Nature*, 414(6865), 799-806.
- Santoro, M. G. (2000). Heat Shock Factors and the Control of the Stress Response. *Biochemical Pharmacology*, 59(1), 55-63.
- Scartezzini, P., & Speroni, E. (2000). Review on Some Plants of Indian Traditional Medicine with Antioxidant Activity. *Journal of Ethnopharmacology*, 71: 23-43.
- Schrijvers, B. F., De Vriese, A. S. and Flyvbjerg, A. (2004). From Hyperglycemia to Diabetic Kidney Disease: The Role of Metabolic, Hemodynamic, Intracellular Factors And Growth Factors/Cytokines. *Endocrine Reviews*, 25(6), 971-1010.
- Scott, K. and Receveur, O. (1995). Ethics for Working with Communities of Indigenous Peoples. *Canadian Journal Of Physiology And Pharmacology*, 73(6), 751-753.
- Seah, R., Siripongvutikorn, S. and Usawakesmanee, W. (2010). Antioxidant and Antibacterial Properties in Keang Hleung Paste and Its Ingredients. *Asian Journal of Food and Agro-Industry*, 3(2), 213-220.
- Sekar, M., bin Abdullah, M. Z., binti Nasir, S. N., binti Zakaria, Z., & bin Abdullah, M. S. (2014). Ten Commonly Available Medicinal Plants in Malaysia Used for the Treatment Of Diabetes—A Review. *Asian Journal of Pharmaceutical and Clinical Research*, 1(2).
- Settharaksa, S., Jongjareonrak, A., Hmadhlu, P., Chansuwan, W. and Siripongvutikorn, S. (2012). Flavonoid, Phenolic Contents and Antioxidant Properties of Thai Hot Curry Paste Extract and Its Ingredients as Affected of pH, Solvent Types and High Temperature. *International Food Research Journal*, 19(4), 1581-1587.
- Seven, A., Guzel, S., Seymen, O., Civelek, S., Bolayirli, M., Uncu, M. (2004). Effects Of Vitamin E Supplementation on Oxidative Stress In Streptozotocin

- Induced Diabetic Rats: Investigation of Liver and Plasma. *Yonsei Medical Journal*, 45(4), 703-710.
- Shahidi, F. and Naczk, M. (1995). *Food Phenolics: Sources, Chemistry, Effects, Applications*: Technomic Publishing Company Lancaster.
- Shaw, J., Sicree, R. and Zimmet, P. (2010). Global Estimates of the Prevalence of Diabetes for 2010 and 2030. *Diabetes Research and Clinical Practice*, 87(1), 4-14.
- Shichijo K, Ihara M, Matsuu M, Ito M, Okumura Y, Sekine I.( 2003). Overexpression of Heat Shock Protein 70 in Stomach of Stress-Induced Gastric Ulcer-Resistant Rats. *Digest Dis Sci*. 48(2) : 340-348.
- Shimabukuro, M., Zhou, Y.-T., Levi, M. and Unger, R. H. (1998). Fatty Acid-Induced  $\beta$  Cell Apoptosis: A Link Between Obesity and Diabetes. *Proceedings of the National Academy of Sciences*, 95(5), 2498-2502.
- Shimada, K., Fujikawa, K., Yahara, K. and Nakamura, T. (1992). Antioxidative Properties of Xanthan on the Autoxidation of Soybean Oil in Cyclodextrin Emulsion. *Journal of Agricultural and Food Chemistry*, 40(6), 945-948.
- Shimoji, Y., Tamura, Y., Nakamura, Y., Nanda, K., Nishidai, S., Nishikawa, Y. (2002). Isolation and Identification of DPPH Radical Scavenging Compounds in Kurosu (Japanese Unpolished Rice Vinegar). *Journal of Agricultural and Food Chemistry*, 50(22), 6501-6503.
- Shobana, S. and Akhilender Naidu, K. (2000). Antioxidant activity of selected Indian Spices. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 62(2), 107-110.
- Siegel, R. D., Cupples, A., Schaefer, E. J. and Wilson, P. W. (1996). Lipoproteins, Apolipoproteins, and Low-Density Lipoprotein Size Among Diabetics in the Framingham Offspring Study. *Metabolism*, 45(10), 1267-1272.
- Sies, H. and Stahl, W. (1995). Vitamins E and C, Beta-Carotene, and Other Carotenoids as Antioxidants. *The American journal of clinical nutrition*, 62(6), 1315S-1321S.
- Simkó, M., Gazsó, A., Fiedeler, U. and Nentwich, M. (2011). Nanoparticles, Free Radicals and Oxidative Stress. *NanoTrust Dossiers*, 12, 1-3.
- Soa, O., A. E. A. Oaa and A. Dab. (2011). Antidiabetic and Haematological Effect of Aqueous Extract of Stem Bark of *Azela africana* (Smith) on Streptozotocin–

- Induced Diabetic Wistar rats. *Asian Pacific journal of tropical biomedicine*, 1(5), 353-358.
- Soto, C., Muriel, P. and Reyes, J. L. (1994). Pancreatic Lipid Peroxidation in Alloxan-Induced Diabetes Mellitus. *Archives of Medical Research*, 25, 377-377.
- Spitaler, M. and Graier, W. (2002). Vascular Targets of Redox Signalling in Diabetes Mellitus. *Diabetologia*, 45(4), 476-494.
- Sriplang, K., Adisakwattana, S., Rungsipipat, A. and Yibchok-anun, S (2007). Effects of Orthosiphon Stamineus Aqueous Extract on Plasma Glucose Concentration and lipid Profile in Normal and Streptozotocin-Induced Diabetic rats. *Journal of Ethnopharmacology*, 109(3), 510-514.
- Stacchiotti, A., Ricci, F., Rezzani, R., Volti, G. L., Borsani, E., Lavazza, A., ... & Rodell, L. F. (2006). Tubular Stress Proteins and Nitric Oxide Synthase Expression in Rat Kidney Exposed to Mercuric Chloride and Melatonin. *Journal of Histochemistry & Cytochemistry*, 54(10), 1149-1157.
- Stadler, K., Jenei, V., von Bölcsházy, G., Somogyi, A. and Jakus, J. (2004). Role of Free Radicals and Reactive Nitrogen Species in the Late Complications of Diabetes Mellitus in Rats. *Orvosi hetilap*, 145(21), 1135.
- Stahl, W., Heinrich, U., Jungmann, H., Sies, H. and Tronnier, H. (2000). Carotenoids and Carotenoids Plus Vitamin E Protect Against Ultraviolet Light-Induced Erythema in Humans. *The American journal of clinical nutrition*, 71(3), 795-798.
- Steenhuysen, J. (2007). Mother Nature Still a Rich Source of New Drugs. *Environmental News Networks [online] Available:(<http://www.enn.com/med.html?idH442>)(April 15, 2007).*
- Stevens, R. B., Sutherland, D., Ansite, J. D., Saxena, M., Rossini, T., Levay-Young, B.(1997). Insulin Down-Regulates the Inducible Nitric Oxide Synthase Pathway: Nitric Oxide as Cause and Effect of Diabetes? *The Journal of Immunology*, 159(11), 5329-5335.
- Stoscheck, CM. (1990). Quantitation of Protein. *Methods in Enzymology* 182: 50-69
- Sumaryono, W.; Proksch, P.; Wray, V.; Witte, L.( 1991). Hartmann, T. Qualitative and Quantitative Analysis of the Phenolic Constituents from Orthosiphon Aristatus. *Planta Med.* 57, 176–180.

- Sunmonu, T. O., & Afolayan, A. J. (2013). Evaluation of Antidiabetic Activity and Associated Toxicity of *Artemisia Afra* Aqueous Extract in Wistar Rats. *Evidence-Based Complementary and Alternative Medicine* . vol. 2013, Article ID 929074, 8 pages, 2013. doi:10.1155/2013/929074.
- Suvarna, K. S., Suvarna, S. K., Layton, C., & Bancroft, J. D. (Eds.). (2012). *Bancroft's Theory and Practice of Histological Techniques*, Expert Consult: Online and Print, 7: *Bancroft's Theory and Practice of Histological Techniques*. Elsevier Health Sciences.
- Suzuki, K., Jayasena, C. N., & Bloom, S. R. (2012). Obesity and Appetite Control. *Experimental Diabetes Research*, 2012.
- Szkudelski, T. (2001). The Mechanism of Alloxan and Streptozotocin Action in B Cells of the Rat Pancreas. *Physiological Research*, 50(6), 537-546.
- Taga, M. S., Miller, E. and Pratt, D. (1984). Chia Seeds as a Source of Natural Lipid Antioxidants. *Journal of the American Oil Chemists' Society*, 61(5), 928-931.
- Taha, A. M.(2002). Traditional/complementary Medicine in the Malaysian Healthcare System. Proceedings of the 2002 *4th International Conference for Traditional/Complementary Medicine*, Sunway Pyramid Convention Centre, Selangor, Malaysia.
- Takada, J., Fonseca-Alaniz, M. H., de Campos, T. B. F., Andreotti, S., Campana, A. B., Okamoto, M., ... & Lima, F. B. (2008). Metabolic recovery of adipose Tissue is Associated with Improvement in insulin Resistance in a Model of Experimental Diabetes. *Journal of Endocrinology*, 198(1), 51-60.
- Talubmook, C. (2008). Effect of Polysaccharide from *Phellinus Ignarius* (L) Quel. on Hematological Values and Blood Cell Characteristic in Diabetic Rats. *Journal of Microscopy Society of Thailand*, 22, 42-45.
- Talukder, M. E. U., Aklima, J., Emran, T. B., Islam, S., Rahman, A., & Bhuiyan, R. H. (2013). In vitro Antioxidant Potential of *Momordica charantia* Fruit Extracts. *British Journal of Pharmaceutical Research*, 3(4), 963-971.
- Tan MH, MacLean DR, (1995). Epidemiology of Diabetes Mellitus in Canada. *Clin Invest Med* 18: 240–246.
- Taskinen, MR.(1992). Quantitative and Qualitative Lipoprotein Abnormalities in Diabetes Mellitus. *Diabetes*, 41(Supplement 2), 12-17.
- Thomas, N., Alder, E. and Leese, G.(2004). Barriers to Physical Activity in Patients with Diabetes. *Postgraduate Medical Journal*, 80(943), 287-291.

- Tiedge, M., Lortz, S., Drinkgern, J. and Lenzen, S.(1997). Relation between Antioxidant Enzyme Gene Expression and Antioxidative Defense Status of Insulin-Producing Cells. *Diabetes*, 46(11), 1733-1742.
- Tierney, L. M., McGhee, S. J., Papadakos, M.A. (2002). *Current Medical Diagnosis and Treatment International Edition*, New York .Large Medical Book, 70, pp. 120-125.
- Tiwari, A. K. and Madhusudanarao, J.(2002). Diabetes Mellitus and Multiple Therapeutic Approaches of Phytochemicals: Present Status and Future Prospects. *Current Science*, 83.
- Twaij, H. A. and Al-Badr, A. A. (1988). Hypoglycemic Activity of *Artemisia herba alba*. *Journal of Ethnopharmacology*, 24(2), 123-126.
- Unger, R. and Grundy, S.(1985). Hyperglycaemia as an Inducer as well as a Consequence of Impaired Islet Cell Function and Insulin Resistance: Implications for the Management of Diabetes. *Diabetologia*, 28(3), 119-121.
- Unger, R. H.(1995). Lipotoxicity in the Pathogenesis of Obesity-dependent NIDDM: Genetic and Clinical Implications. *Diabetes*, 44(8), 863-870.
- Velioglu, Y., Mazza, G., Gao, L. and Oomah, B.(1998). Antioxidant Activity and Total Phenolics in Selected Fruits, Vegetables, and Grain Products. *Journal of Agricultural and Food Chemistry*, 46(10), 4113-4117.
- Venkat Narayan, K., Gregg, E. W., Fagot-Campagna, A., Engelgau, M. M. and Vinicor, F. (2000). Diabetes—A Common, Growing, Serious, Costly, and Potentially Preventable Public Health Problem. *Diabetes Research and Clinical Practice*, 50, S77-S84.
- Verma, S. and Singh, S. (2008). Current and Future Status of Herbal Medicines. *Veterinary World*, 1(11), 347-350.
- Vessal, M., Hemmati, M. and Vasei, M. (2003). Antidiabetic Effects of Quercetin in Streptozocin-induced Diabetic Rats. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 135(3), 357-364.
- Vinicor, F. (1994). Is Diabetes a Public-health Disorder? *Diabetes Care*, 17, 22.
- Vinicor, F.(1998). The Public Health Burden of Diabetes and the Reality of Limits. *Diabetes Care*, 21(Supplement 3), C15-C18.
- Voegeli, T. S., Wintink, A. J., Chen, Y. and Currie, R. W.(2008). Heat Shock Proteins 27 and 70 Regulating Angiotensin II-Induced NF- $\kappa$  B: A Possible



- Connection to Blood Pressure Control? *Applied Physiology, Nutrition, and Metabolism*, 33(5), 1042-1049.
- Vrekoussis, T., Chaniotis, V., Navrozoglou, I., Dousias, V., Pavlakis, K., Stathopoulos, E. N., & Zoras, O. (2009). Image Analysis of Breast Cancer Immunohistochemistry-Stained Sections using ImageJ: an RGB-based Model. *Anticancer Research*, 29(12), 4995-4998.
- Walzem, R., Watkins, S., Frankel, E., Hansen, R. and German, J.(1995). Older Plasma Lipoproteins are More Susceptible to Oxidation: A Linking Mechanism for the Lipid and Oxidation Theories of Atherosclerotic Cardiovascular Disease. *Proceedings of the National Academy of Sciences*, 92(16), 7460-7464.
- Wang, Z. and Gleichmann, H.(1998). GLUT2 in Pancreatic Islets: Crucial Target Molecule in Diabetes Induced with Multiple Low Doses of Streptozotocin in Mice. *Diabetes*, 47(1), 50-56.
- Weinberg, J. B.(2000). Nitric Oxide Synthase 2 and Cyclooxygenase 2 Interactions in Inflammation. *Immunologic Research*, 22(2-3), 319-341.
- Wei W, Liu Q, Tan Y, Liu L, Li X, Cai L.( 2009). Oxidative stress, diabetes, and diabetic complications. *Hemoglobin*. ;33:370–377.
- West, K. M., Ahuja, M., Bennett, P. H., Czyzyk, A., De Acosta, O. M., Fuller, J. H. (1983). The Role of Circulating Glucose and Triglyceride Concentrations and Their Interactions with Other “Risk Factors” as Determinants of Arterial Disease in Nine Diabetic Population Samples from the WHO Multinational Study. *Diabetes Care*, 6(4), 361-369.
- Whiting, D. R., Guariguata, L., Weil, C. and Shaw, J. (2011). IDF Diabetes Atlas: Global Estimates of the Prevalence of Diabetes for 2011 and 2030. *Diabetes Research and Clinical Practice*, 94(3), 311-321.
- Williams, L. (2006). Ethnomedicine. *West Indian Medical Journal*, 55(4), 215-216.
- World Health Organization. (1980). *WHO Expert Committee on Diabetes Mellitus: Second Report*, Technical Report Series 646. WHO, Geneva.
- World Health Organization. (1985). *WHO Expert Committee on Diabetes Mellitus: Second Report*, Technical Report Series no. 844. WHO, Geneva
- World Health Organization. (1999). *Definition, Diagnosis and Classification of Diabetes Mellitus and Its Complications*: World Health Organization.

- World Health Organization .(2003). Traditional Medicine Fact Sheet No 134. Retrieved from <http://www.who.int/mediacentre/factsheets/2003/fs134/en/>.
- World Health Organization. (2009a). Traditional medicine Fact sheet No 134. 2003. *Online document at: www. who. int/mediacentre/factsheets/fs134/en/* Accessed May, 5.
- World Health Organization .(1999b). *WHO Monographs on Selected Medicinal Plants Vol. 1. Vol. 1*. Geneva: World Health Organization.
- Yoopan, N., Thisoda, P., Rangkadilok, N., Sahasitiwat, S., Pholphana, N., Ruchirawat,S.(2007). Cardiovascular Effects of 14-deoxy-11, 12-Didehydroandrographolide and Andrographis Paniculata Extracts. *Planta Medica*, 73(06), 503-511.
- Yoshikawa, T.(1997). Science of Free Radicals. Koudan Sha Saientifikku, Tokyo, .
- Yoshikawa, T. and Naito, Y.(2002). What is Oxidative Stress? *Japan Medical Association Journal*, 45(7), 271-276.
- Youn, M. Y.(2012). *The Effect of Intake of Whole Grain or Whole Grain Components on Type 2 Diabetes in Rats*. 3498551 Ph.D., University of Minnesota, United States
- Zajac, J., Shrestha, A., Patel, P. and Poretsky, L.(2010). The Main Events in the History of Diabetes Mellitus. *Principles of Diabetes Mellitus* (pp. 3-16): Springer.
- Zakaria, M. and Mohd, M. A. (1994). *Traditional Malay Medicinal Plants.*: Penerbit Fajar Bakti, Sdn. Bhd., Kuala Lumpur.
- Zeid, I. M. A., (2010). Identification of some Date Palm ( Phoenix dactylifera ) Cultivars by Fruit Characters A B C D E F. *Journal of Science and Technology*, 3: 338-343.
- Zhao, Y., Yang, X., Lu, W., Liao, H., & Liao, F. (2009). Uricase Based Methods for Determination of Uric Acid in Serum. *Microchimica Acta*, 164(1-2), 1-6.
- Zheng, W. and Wang, S. Y. (2001). Antioxidant Activity and Phenolic Compounds in Selected Herbs. *Journal of Agricultural and Food Chemistry*, 49(11), 5165-5170.
- Zhishen, J., Mengcheng, T. and Jianming, W.(1999). The Determination of Flavonoid Contents in Mulberry and Their Scavenging Effects on Superoxide Radicals. *Food Chemistry*, 64(4), 555-559.

Zimmerlin, L., Donnenberg, A. D., Rubin, J. P., Basse, P., Landreneau, R. J., & Donnenberg, V. S. (2010). Regenerative Therapy and Cancer: In vitro and in Vivo Studies of the Interaction between Adipose-Derived Stem Cells and Breast Cancer Cells from Clinical Isolates. *Tissue Engineering Part A*, 17(1-2), 93-106.